

Mouse anti-Human C-Reactive Protein (CRP) ELISA Set

Cat. No.	Form	Quantity
11085-01	ELISA Set	1

For Research Use Only

Intended Use

This set is specifically designed for quantitative determination of human C-reactive protein (CRP) concentrations in cell culture supernatant, plasma and serum.

Background

C-reactive protein (CRP) is a member of the acute phase proteins. CRP levels rise dramatically during inflammatory processes occurring in the body and it is thought to assist in complement binding to foreign and damaged cells and affect the humoral response to disease. Research has shown patients with elevated levels of CRP to be at an increased risk for diabetes, hypertension and cardiovascular disease^{1,2}. High CRP concentrations in plasma may be associated with inflammation that is a risk factor for the development of colon cancer in average-risk individuals³. In addition, CRP may exacerbate ischemic necrosis in a complement-dependent fashion and CRP inhibition may be a safe and effective therapy for myocardial and cerebral infarcts⁴.

Principle of Assay

This assay incorporates a quantitative sandwich enzyme immunoassay technique. Unlabeled monoclonal antibody specific for human CRP is first coated onto 96 well microplate(s). Standards and samples are then added to the wells and any CRP present is captured by the immobilized antibody. After a wash to remove unbound material, HRP-labeled anti-human CRP detection antibody is added to the wells. After another wash, TMB substrate solution is added that will result in a blue color proportional to the amount of bound CRP. Color development is then quenched and intensity is measured at 450nm.

Research Applications

Enzyme-Linked-Immunosorbent-Assay (ELISA)

Characterization

To ensure lot-to-lot consistency, each batch of product is tested by ELISA to conform to the characteristics of a standard reference reagent.

Assay Kit Components

- 0.3mL CRP Capture Antibody: unlabeled, 200X
- 0.3mL CRP Detection Antibody: HRP-conjugated, 200X
- 2 vials Standard: lyophilized recombinant human CRP, 50ng/vial

- 15mL Coating Buffer (5X)
- 300mL Assay Diluent (1X)
- 300mL Washing Buffer (20X)
- 60mL Standard/Sample Diluent (1X)
- 60mL TMB one component microwell substrate
- 60mL TMB Stop Solution
- 5 x 96-well ELISA plates

Sample Collection and Storage

Cell Culture Supernatant - Remove particulates by centrifugation and assay immediately or aliquot and store samples at -20°C. Avoid repeated freeze-thaw cycles.

Plasma - Collect plasma using citrate, EDTA or heparin as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at -20°C. Avoid repeated freeze-thaw cycles.

Serum - Use a serum separator tube and allow samples to clot for 30 minutes before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at -20°C. Avoid repeated freeze-thaw cycles.

Note: Plasma and serum samples require a 200-fold dilution. A suggested 200-fold dilution is 5.0µL sample into 995µL Assay Diluent.

Reagent Preparation

All reagents should be diluted immediately prior to use. Do not add sodium azide to Assay Diluent as it inhibits the activity of horseradish-peroxidase. It is recommended that all samples, controls and standards be assayed in duplicate or triplicate.

- Dilute 5X Coating Buffer to 1X with distilled (DI) water. A suggested dilution (for one plate) consists of 2.2mL 5X Coating Buffer with 8.8mL of DI water.
- Dilute unlabeled human CRP specific Capture Antibody 200-fold with 1X Coating Buffer. A suggested dilution (for one plate) consists of 55µL Capture Antibody with 10.945mL 1X Coating Buffer.
- Dilute 20X Wash Buffer to 1X with DI water. A suggested dilution consists of 50mL 20X Wash Buffer with 950mL of DI water.
- Reconstitute 1 vial lyophilized human CRP Standard by adding 0.1mL DI water to 1 vial (500ng/mL) and gently vortex. Next, dilute CRP Standard 100-fold by adding 5µL 500ng/mL Standard to 495µL DI water to yield a final concentration of 5ng/mL and allow the standard to equilibrate for at least 15 minutes. Aliquot any unused standard into polypropylene vials and store at -80°C.
- Dilute HRP-labeled human CRP specific Detection Antibody 200-fold with 1X Assay Diluent. A suggested dilution (for one plate) consists of 55µL Detection Antibody with 10.945mL 1X Assay Diluent.

Assay Procedure

1. Collect samples and prepare reagents as directed previously.
2. Add 100µL of diluted Capture Antibody solution to all wells of a provided 96-well plate. Incubate at 4°C overnight.
3. Wash entire plate 4 times with 1X Wash Buffer.
4. Add 300µL of 1X Assay Diluent per well to block non-specific binding and reduce background. Incubate at room temperature for 30 minutes.
5. Wash plate as in Step 3.
6. Prepare standard dilution by adding 100µL/well of appropriate dilution to the plate. To do this, perform six two-fold serial dilutions of the 5ng/mL top standard either within the plate or in separate tubes. To dilute within the plate, add 200µL of 5ng/mL human CRP Standard to row A and 100µL Standard/Sample Diluent to rows B through H. Perform serial dilution by taking 100µL standard from row A and mix with row B. Then take 100µL from row B and add it to row C and continue for each subsequent row. Therefore, the final human CRP standard concentrations will be 5ng/mL, 2.5ng/mL, 1.25ng/mL, 0.63ng/mL, 0.31ng/mL, 0.16ng/mL and 0.08ng/mL. Standard/Sample Diluent alone serves as the zero standards for row H (0ng/mL). For test samples, dilute two-fold with Standard/Sample Diluent and add 100µL/well. Incubate at room temperature for 2 hours.
7. Wash plate as in Step 3.
8. Add 100µL of diluted HRP-labeled Detection Antibody solution to each well and incubate at room temperature for 1 hour.
9. Wash plate 6 times with 1X Wash Buffer. For this final wash, soak wells in Wash Buffer for 10 seconds for each wash. This will help minimize background.
10. Add 100µL of TMB Substrate Solution to each well. Incubate at room temperature for 10-15 minutes. The Substrate Solution should turn blue in color.
11. Add 50µL of Stop Solution to each well. The color in the wells should change from blue to yellow. If the color in the wells turns green or the color change does not appear uniform, gently tap the plate to ensure thorough mixing.
12. Determine the optical density of each well within 30 minutes using a microplate reader set to 450 nm.

Specificity

This assay recognizes recombinant and native human CRP. No cross-reactivity or interference with other human proteins was observed.

Sensitivity

The minimum detectable concentration of human CRP is 0.045ng/mL.

Recovery

The recovery of human CRP spiked to levels throughout the range of the assay in cell culture supernatant was evaluated by the set. At least 90% of the CRP was recovered from cell culture supernatant.

Trouble Shooting

General technical hints:

- To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions and between reagent additions. Also, use separate reservoirs for each reagent.
- When using an automated plate washer, adding a 30 second soak period following the addition of wash buffer and/or rotating the plate 180 degrees between wash steps may improve assay precision.
- During incubation steps, shaking the plates may increase sensitivity.
- Substrate Solution should remain colorless until added to the plate. Keep Substrate Solution protected from light. Substrate Solution should change from colorless to gradations of blue upon addition.
- Stop Solution should be added to the plate in the same order as the Substrate Solution.
- The color development in the wells will turn from blue to yellow upon addition of the Stop Solution. Wells that turn green in color indicate that the Stop Solution has not mixed thoroughly with the Substrate Solution.

Poor Signal

- Avoid sodium azide in wash buffers as it inhibits the enzymatic activity of HRP
- Verify that appropriate antibody pairs were used
- Inadequate reagent volumes added to wells
- Incorrect incubation times and/or temperature

Poor Standard Curve or Precision

- Improper standard handling and/or dilution
- Inadequate mixing of reagents
- Inadequate aspiration and/or washing of wells

References

1. Torelli, Julius. (2005). Beyond Cholesterol, p.45. ISBN 0-312-34863-0.
2. Pradhan, AD. (2001). *JAMA*. **286**: 327-34.
3. Erlinger, TP, Platz, EA, Rifai, N, Helzlsouer KJ (2004). *JAMA*. **291**: 585-90.
4. Pepys, MB et al. (2006). *Nature* **440**: 1217-21.