



Rat Anti-Mouse IL-2

Cat. No.	Format	Size
10201-01	Purified (UNLB)	0.5 mg
10201-08	Biotin (BIOT)	0.5 mg
10201-14	Low Endotoxin, Azide-Free (LE/AF)	0.5 mg

Overview

Clone JES6-1A12 Isotype Rat IgG_{2a}κ

E. coli-expressed mouse IL-2 Immunogen

Specificity Mouse IL-2

Alternate Name(s) Interleukin-2, thymocyte differentiation factor, TDF, T cell growth factor, TCGF, killer cell helper factor,

KHF, macrophage-activating factor for cytotoxicity I, MAF-C I, eosinophil differentiation factor, EDF

Applications

ELISA-Capture - Quality tested 1-6 ELISA-Detection - Quality tested ELISPOT-Capture - Reported in literature 5,7-14 WB - Reported in literature 15 IP - Reported in literature 1 Neut – Reported in literature 1,2,16 Multiplex-Capture - Reported in literature 3

Working Dilutions

ELISA Purified (UNLB) antibody ≤ 5 μg/mL

> BIOT conjugate 1:1,000 - 1:4,000

Since applications vary, you should determine the optimum working dilution for the product that is Other Applications

appropriate for your specific need.

For Research Use Only. Not for Diagnostic or Therapeutic Use.

Handling and Storage

- The purified (UNLB) antibody is supplied as 0.5 mg purified immunoglobulin in 1.0 mL of borate buffered saline, pH 8.2. No preservatives or amine-containing buffer salts added. Store at 2-8°C.
- The biotin (BIOT) conjugate is supplied as 0.5 mg labeled antibody in 1.0 mL of PBS/NaN₃. Store at 2-8°C.
- The low endotoxin, azide-free (LE/AF) antibody is supplied as 0.5 mg purified immunoglobulin in 1.0 mL of PBS. Contains no
 preservative; handle under aseptic conditions. Store at 2-8°C or aliquot into smaller volumes and store at -20°C. Avoid multiple
 freeze / thaw cycles.
- Reagents are stable for the period shown on the label if stored as directed.

Warning

Some reagents contain sodium azide. Please refer to product specific SDS.

References

- Sander B, Höidén I, Andersson U, Möller E, Abrams JS. Similar frequencies and kinetics of cytokine producing cells in murine peripheral blood and spleen. Cytokine detection by immunoassay and intracellular immunostaining. J Immunol Methods. 1993;166:201-14. (Immunogen, ELISA-Capture, IP, Neut)
- Abrams JS, Roncarolo M, Yssel H, Andersson U, Gleich GJ, Silver JE. Strategies of anti-cytokine monoclonal antibody development: immunoassay of IL-10 and IL-5 in clinical samples. Immunol Rev. 1992;127:5-24. (ELISA-Capture, Neut)
- Carson RT, Vignali DA. Simultaneous quantitation of 15 cytokines using a multiplexed flow cytometric assay. J Immunol Methods. 1999;227:41-52. (ELISA-Capture, Multiplex-Capture)
- Abrams JS. Immunoenzymetric assay of mouse and human cytokines using NIP-labeled anti-cytokine antibodies. Curr Protoc Immunol. 2001;6.20:1-15. (ELISA-Capture)
- Pack CD, Cestra AE, Min B, Legge KL, Li L, Caprio-Young JC, et al. Neonatal exposure to antigen primes the immune system to develop responses in various lymphoid organs and promotes bystander regulation of diverse T cell specificities. J Immunol. 2001;167:4187-95. (ELISA-Capture, ELISPOT-Capture)
- 6. Gessner A, Mohrs K, Mohrs M. Mast cells, basophils, and eosinophils acquire constitutive IL-4 and IL-13 transcripts during lineage differentiation that are sufficient for rapid cytokine production. J Immunol. 2005;174:1063-72. (ELISA-Capture)
- 7. Klinman D. ELISPOT assay to detect cytokine-secreting murine and human cells. Curr Protoc Immunol. 2008;6.19:1-9. (ELISPOT-Capture)
- 8. Karulin AY, Hesse MD, Tary-Lehmann M, Lehmann PV. Single-cytokine-producing CD4 memory cells predominate in type 1 and type 2 immunity. J Immunol. 2000;164:1862-72. (ELISPOT-Capture)
- 9. Fedoseyeva EV, Kishimoto K, Rolls HK, Illigens BM, Dong VM, Valujskikh A, et al. Modulation of tissue-specific immune response to cardiac myosin can prolong survival of allogeneic heart transplants. J Immunol. 2002;169:1168-74. (ELISPOT-Capture)
- Stern BV, Boehm BO, Tary-Lehmann M. Vaccination with tumor peptide in CpG adjuvant protects via IFN-γ-dependent CD4 cell immunity. J Immunol. 2002;168:6099-105. (ELISPOT-Capture)
- 11. Kreher CR, Dittrich MT, Guerkov R, Boehm BO, Tary-Lehmann M. CD4⁺ and CD8⁺ cells in cryopreserved human PBMC maintain full functionality in cytokine ELISPOT assays. J Immunol Methods. 2003;278:79-93. (ELISPOT-Capture)
- 12. Nekrasova T, Shive C, Gao Y, Kawamura K, Guardia R, Landreth G, et al. ERK1-deficient mice show normal T cell effector function and are highly susceptible to experimental autoimmune encephalomyelitis. J Immunol. 2005;175:2374-80. (ELISPOT-Capture)
- 13. Linker RA, Rott E, Hofstetter HH, Hanke T, Toyka KV, Gold R. EAE in beta-2 microglobulin-deficient mice: axonal damage is not dependent on MHC-I restricted immune responses. Neurobiol Dis. 2005;19:218-28. (ELISPOT-Capture)
- 14. Hofstetter HH, Mössner R, Lesch KP, Linker RA, Toyka KV, Gold R. Absence of reuptake of serotonin influences susceptibility to clinical autoimmune disease and neuroantigen-specific interferon-gamma production in mouse EAE. Clin Exp Immunol. 2005;142:39-44. (ELISPOT-Capture)
- 15. Liu S, Sher Y, Ting C, Liao K, Yu C, Tao M. Treatment of B-cell lymphoma with chimeric IgG and single-chain Fv antibody-interleukin-2 fusion proteins. Blood. 1998;92:2103-12. (WB)
- 16. van Hamburg JP, de Bruijn MJ, de Almeida CR, van Zwam M, van Meurs M, de Haas E, et al. Enforced expression of GATA3 allows differentiation of IL-17-producing cells, but constrains Th17-mediated pathology. Eur J Immunol. 2008;38:2573-86. (Neut)

TB10201 08-Oct-21