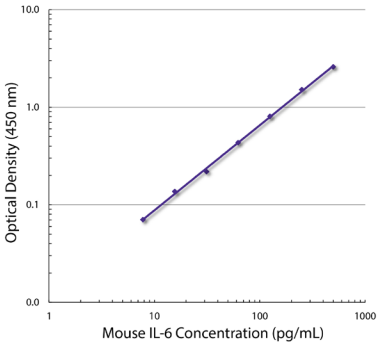
SouthernBiotech

Rat Anti-Mouse IL-6

Cat. No.	Format	Size
10207-01	Purified (UNLB)	0.5 mg
10207-14	Low Endotoxin, Azide-Free (LE/AF)	0.5 mg



Standard curve generated with Rat Anti-Mouse IL-6-UNLB (SB Cat. No. 10207-01; Clone MP5-20F3) and Rat Anti-Mouse IL-6-BIOT (SB Cat. No. 10208-08; Clone MP5-32C11) followed by Mouse Anti-BIOT-HRP (SB Cat. No. 6404-05)

Overview

Clone MP5-20F3
Isotype Rat $IgG_1\kappa$
Immunogen COS-expressed mouse IL-6
Specificity Mouse IL-6
Alternate Name(s) Interleukin-6, hepatocyte stimulating factor, HSF, hybridoma/plasmacytoma growth factor, HPGFB cell stimulating factor-2, BSF-2, cytotoxic T cell differentiation factor, CDF

Applications

ELISA-Capture – Quality tested [1-7]
ELISPOT-Capture – Reported in literature [4,8-10]
FC – Reported in literature [11-14]
IHC-FS – Reported in literature [15-19]
IHC-PS – Reported in literature [20-22]
ICC – Reported in literature [3]
IP – Reported in literature [1]
Neut – Reported in literature [1-3,23]
Multiplex-Capture – Reported in literature [5]

Note – May be paired with the biotinylated clone MP5-32C11 (SB Cat. No. 10208-08) in a sandwich ELISA

Working Dilutions

ELISA	Purified (UNLB) antibody	≤ 2 μg/mL
Other Applications	Since applications vary, you should determine the optimum working dilution for the product that is appropriate for your specific need.	

For Research Use Only. Not for Diagnostic or Therapeutic Use.

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Handling and Storage

- The purified (UNLB) antibody is supplied as 0.5 mg purified immunoglobulin in 1.0 mL of borate buffered saline, pH 8.2. *No preservatives or amine-containing buffer salts added*. Store at 2-8°C.
- The low endotoxin, azide-free (LE/AF) antibody is supplied as 0.5 mg purified immunoglobulin in 1.0 mL of PBS. Contains no preservative; handle under aseptic conditions. Store at 2-8°C or aliquot into smaller volumes and store at -20°C. Avoid multiple freeze / thaw cycles.
- Reagents are stable for the period shown on the label if stored as directed.

References

1. Starnes HF Jr, Pearce MK, Tewari A, Yim JH, Zou J, Abrams JS. Anti-IL-6 monoclonal antibodies protect against lethal Escherichia coli infection and lethal tumor necrosis factor-α challenge in mice. J Immunol. 1990;145:4185-91. (Immunogen, ELISA-Capture, IP, Neut)
2. Abrams JS, Roncarolo M, Yssel H, Andersson U, Gleich GJ, Silver JE. Strategies of anti-cytokine monoclonal antibody development: immunoassay of IL-10 and IL-5 in clinical samples. Immunol Rev. 1992;127:5-24. (ELISA-Capture, Neut)
3. Sander B, Höidén I, Andersson U, Möller E, Abrams JS. Similar frequencies and kinetics of cytokine producing cells in murine peripheral blood and spleen. Cytokine detection by immunoassay and intracellular immunostaining. J Immunol Methods. 1993;166:201-14. (ELISA-Capture, ICC, Neut)
4. Shirai A, Holmes K, Klinman D. Detection and quantitation of cells secreting IL-6 under physiologic conditions in BALB/c mice. J Immunol. 1993;150:793-9. (ELISA-Capture, ELISPOT-Capture)
5. Carson RT, Vignali DA. Simultaneous quantitation of 15 cytokines using a multiplexed flow cytometric assay. J Immunol Methods. 1999;227:41-52. (ELISA-Capture, Multiplex-Capture)
6. da Fonseca DM, Silva CL, Wowk PF, Paula MO, Ramos SG, Horn C, et al. Mycobacterium tuberculosis culture filtrate proteins plus CpG Oligodeoxynucleotides confer protection to Mycobacterium bovis BCG-primed mice by inhibiting interleukin-4 secretion. Infect Immun. 2009;77:5311-21. (ELISA-Capture)
7. Abrams JS. Immunoenzymetric assay of mouse and human cytokines using NIP-labeled anti-cytokine antibodies. Curr Protoc Immunol. 2001;6.20:1-15. (ELISA-Capture)
8. Stern BV, Boehm BO, Tary-Lehmann M. Vaccination with tumor peptide in CpG adjuvant protects via IFN-γ-dependent CD4 cell immunity. J Immunol. 2002;168:6099-105. (ELISPOT-Capture)
9. Linker RA, Rott E, Hofstetter HH, Hanke T, Toyka KV, Gold R. EAE in beta-2 microglobulin-deficient mice: axonal damage is not dependent on MHC-I restricted immune responses. Neurobiol Dis. 2005;19:218-28. (ELISPOT-Capture)
10. Klinman D. ELISPOT assay to detect cytokine-secreting murine and human cells. Curr Protoc Immunol. 2008;6.19:1-9. (ELISPOT-Capture)
11. Tripp RA, Moore D, Jones L, Sullender W, Winter J, Anderson LJ. Respiratory syncytial virus G and/or SH protein alters Th1 cytokines, natural killer cells, and neutrophils responding to pulmonary infection in BALB/c mice. J Virol. 1999;73:7099-107. (FC)
12. Feng H, Zhang D, Palliser D, Zhu P, Cai S, Schlesinger A, et al. Listeria-infected myeloid dendritic cells produce IFN-β, priming T cell activation. J Immunol. 2005;175:421-32. (FC)
13. Le Huu D, Matsushita T, Jin G, Hamaguchi Y, Hasegawa M, Takehara K, et al. Donor-derived regulatory B cells are important for suppression of murine sclerodermatous chronic graft-versus-host disease. Blood. 2013;121:3274-83. (FC)
14. Nakamura T, Nakao T, Yoshimura N, Ashihara E. Rapamycin prolongs cardiac allograft survival in a mouse model by inducing myeloid-derived suppressor cells. Am J Transplant. 2015 May 5. doi: 10.1111/ajt.13276. [Epub ahead of print]. (FC)
15. Sunnemark D, Ulfgren A, Örn A, Harris RA. Cytokine production in hearts of Trypanosoma cruzi-infected CBA mice: Do cytokine patterns in chronic stage reflect the establishment of myocardial pathology?. Scand J Immunol. 1996;44:421-9. (IHC-FS)
16. Morris MM, Dyson H, Baker D, Harbige LS, Fazakerley JK, Amor S. Characterization of the cellular and cytokine response in the central nervous system following Semliki Forest virus infection. J Neuroimmunol. 1997;74:185-97. (IHC-FS)
17. Marinova-Mutafchieva L, Williams RO, Mason LJ, Mauri C, Feldmann M, Maini RN. Dynamics of proinflammatory cytokine expression in the joints of mice with collagen-induced arthritis (CIA). Clin Exp Immunol. 1997;107:507-12. (IHC-FS)
18. Hersmann GH, Kriegsmann J, Simon J, Hüttich C, Bräuer R. Expression of cell adhesion molecules and cytokines in murine antigen-induced arthritis. Cell Adhes Commun. 1998;6:69-82. (IHC-FS)
19. Schön MP, Schön M, Warren HB, Donohue JP, Parker CM. Cutaneous inflammatory disorder in integrin α_E (CD103)-deficient mice. J Immunol. 2000;165:6583-9. (IHC-FS)
20. Romero-Trevejo JL, Gómez-Villamandos JC, Pedrera M, Blanco A, Bautista MJ, Sánchez-Cordón PJ. Immunohistochemical study of macrophage and cytokine dynamics in the gut of scrapie-infected mice. Histol Histopathol. 2010;25:1025-38. (IHC-PS)
21. Huang T, Tsai S, Liu L, Liu YL, Liu H, Chuang KP. Effect of Arctium lappa L. in the dextran sulfate sodium colitis mouse model. World J Gastroenterol. 2010;16:4193-9. (IHC-PS)
22. Whiteland JL, Shimeld C, Nicholls SM, Easty DL, Williams NA, Hill TJ. Immunohistochemical detection of cytokines in paraffin-embedded mouse tissues. J Immunol Methods. 1997;210:103-8. (IHC-PS)
23. West DM, Del Rosso CR, Yin X, Stuart PM. CXCL1 but not IL-6 is required for recurrent herpetic stromal keratitis. J Immunol. 2014;192:1762-7. (Neut)

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