



## Rat Anti-Mouse IL-17A

Cat. No.	Format	Size
10215-01	Purified (UNLB)	0.5 mg
10215-14	Low Endotoxin, Azide-Free (LE/AF)	0.5 mg

### Overview

<b>Clone</b>	TC11-18H10
<b>Isotype</b>	Rat IgG <sub>1</sub> K
<b>Immunogen</b>	<i>E. coli</i> -expressed mouse IL-17
<b>Specificity</b>	Mouse IL-17A
<b>Alternate Name(s)</b>	Interleukin-17A, cytotoxic T lymphocyte-associated antigen 8, CTLA-8

### Applications

ELISA-Capture – Quality tested <sup>1</sup>  
 ELISPOT-Capture – Reported in literature <sup>2,3</sup>  
 FC – Reported in literature <sup>4,5</sup>  
 Neut – Reported in literature <sup>6-12</sup>

Note – May be paired with the biotinylated clone TC11-8H4 (SB Cat. No. 10214-08) in a sandwich ELISA

### Working Dilutions

<b>ELISA</b>	Purified (UNLB) antibody	≤ 2 µg/mL
<b>Other Applications</b>	Since applications vary, you should determine the optimum working dilution for the product that is appropriate for your specific need.	

***For Research Use Only. Not for Diagnostic or Therapeutic Use.***

## Handling and Storage

---

- The purified (UNLB) antibody is supplied as 0.5 mg purified immunoglobulin in 1.0 mL of borate buffered saline, pH 8.2. *No preservatives or amine-containing buffer salts added.* Store at 2-8°C.
- The low endotoxin, azide-free (LE/AF) antibody is supplied as 0.5 mg purified immunoglobulin in 1.0 mL of PBS. Contains no preservative; handle under aseptic conditions. Store at 2-8°C or aliquot into smaller volumes and store at -20°C. Avoid multiple freeze / thaw cycles.
- Reagents are stable for the period shown on the label if stored as directed.

## References

---

1. Amsen D, de Visser KE, Town T. Approaches to determine expression of inflammatory cytokines. *Methods Mol Biol.* 2009;511:107-42. (ELISA-Capture)
2. Nekrasova T, Shive C, Gao Y, Kawamura K, Guardia R, Landreth G, et al. ERK1-deficient mice show normal T cell effector function and are highly susceptible to experimental autoimmune encephalomyelitis. *J Immunol.* 2005;175:2374-80. (ELISPOT-Capture)
3. Faust SM, Lu G, Marini BL, Zou W, Gordon D, Iwakura Y, et al. Role of T cell TGF $\beta$  signaling and IL-17 in allograft acceptance and fibrosis associated with chronic rejection. *J Immunol.* 2009;183:7297-306. (ELISPOT-Capture)
4. Hamada H, Garcia-Hernandez Mde L, Reome JB, Misra SK, Strutt TM, McKinstry KK, et al. Tc17, a unique subset of CD8 T cells that can protect against lethal influenza challenge. *J Immunol.* 2009;182:3469-81. (FC)
5. Le Huu D, Matsushita T, Jin G, Hamaguchi Y, Hasegawa M, Takehara K, et al. Donor-derived regulatory B cells are important for suppression of murine sclerodermatous chronic graft-versus-host disease. *Blood.* 2013;121:3274-83. (FC)
6. He D, Wu L, Kim HK, Li H, Elmets CA, Xu H. CD8<sup>+</sup> IL-17-producing T cells are important in effector functions for the elicitation of contact hypersensitivity responses. *J Immunol.* 2006;177:6852-8. (Neut)
7. Yusuf N, Nasti TH, Long JA, Naseemuddin M, Lucas AP, Xu H, et al. Protective role of Toll-like receptor 4 during the initiation stage of cutaneous chemical carcinogenesis. *Cancer Res.* 2008;68:615-22. (Neut)
8. He D, Wu L, Kim HK, Li H, Elmets CA, Xu H. IL-17 and IFN- $\gamma$  mediate the elicitation of contact hypersensitivity responses by different mechanisms and both are required for optimal responses. *J Immunol.* 2009;183:1463-70. (Neut)
9. Xiao M, Wang C, Zhang J, Li Z, Zhao X, Qin Z. IFN $\gamma$  promotes papilloma development by up-regulating Th17-associated inflammation. *Cancer Res.* 2009;69:2010-7. (Neut)
10. He D, Li H, Yusuf N, Elmets CA, Li J, Mountz JD, et al. IL-17 promotes tumor development through the induction of tumor promoting microenvironments at tumor sites and myeloid-derived suppressor cells. *J Immunol.* 2010;184:2281-8. (Neut)
11. Kish DD, Volokh N, Baldwin WM 3<sup>rd</sup>, Fairchild RL. Hapten application to the skin induces an inflammatory program directing hapten-primed effector CD8 T cell interaction with hapten-presenting endothelial cells. *J Immunol.* 2011;186:2117-26. (Neut)
12. Karmakar S, Bhaumik SK, Paul J, De T. TLR4 and NKT cell synergy in immunotherapy against visceral leishmaniasis. *PLoS Pathog.* 2012;8(4):e1002646. (Neut)