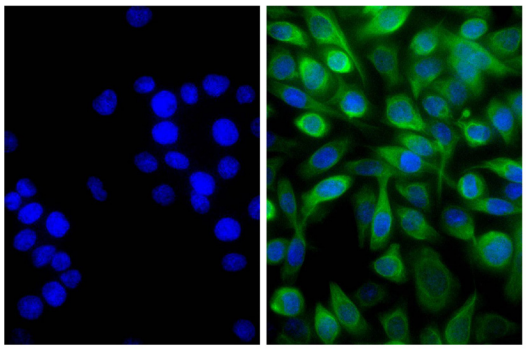
SouthernBiotech

Goat F(ab')$_2$ Anti-Mouse IgG(H+L), Human ads

Cat. No.	Format	Size
1032-01	Purified (UNLB)	0.5 mg
1032-02	Fluorescein (FITC)	0.5 mg
1032-04	Alkaline Phosphatase (AP)	1.0 mL
1032-05	Horseradish Peroxidase (HRP)	1.0 mL
1032-08	Biotin (BIOT)	0.5 mg
1032-09	R-phycoerythrin (PE)	0.25 mg
1032-11	Allophycocyanin (APC)	0.25 mg
1032-13	Spectral Red® (SPRD)	0.25 mg
1032-15	Cyanine 5 (CY5)	0.5 mg
1032-16	R-phycoerythrin-Cyanine 5.5 (PE/CY5.5)	0.25 mg
1032-30	Alexa Fluor® 488 (AF488)	0.5 mg
1032-32	Alexa Fluor® 555 (AF555)	0.5 mg



Human pancreatic carcinoma cell line MIA PaCa-2 was stained with Mouse Anti-Cytokeratin 18-UNLB (SB Cat. No. 10085-01; right) followed by Goat F(ab')2 Anti-Mouse IgG(H+L), Human ads-AF488 (SB Cat. No. 1032-30) and DAPI.

Description

Specificity Reacts with the heavy and light chains of mouse IgG_1, IgG_{2a}, IgG_{2b}, IgG_{2c}, and IgG_3 and with the light chains of mouse IgM and IgA

Source Pepsin digest of Goat Anti-Mouse IgG(H+L), Human ads (SB Cat. No. 1031)

Cross Adsorption Human immunoglobulins and pooled sera; may react with immunoglobulins from other species

Applications

Quality tested applications include –

ELISA [1,2,24,25]
FLISA
FC [4-20]

Other referenced applications include –

ELISPOT [3]
IHC-FS [21,26,27]
IHC-PS [21]
ICC [13,17,22]
WB [23,28]

Working Dilutions

ELISA	AP conjugate	1:2,000 – 1:4,000
	HRP conjugate	1:4,000 – 1:8,000
	BIOT conjugate	1:5,000 – 1:20,000
FLISA	FITC and AF488 conjugates	1:100 – 1:400
	AF555 conjugate	1:50 – 1:200
	PE, APC, and CY5 conjugates	≤ 1 µg/mL
Flow Cytometry	FITC, BIOT, and AF488 conjugates	≤ 1 µg/10^6 cells
	PE, APC, SPRD, and CY5 conjugates	≤ 0.1 µg/10^6 cells
	PE/CY5.5 conjugate	≤ 0.05 µg/10^6 cells
	For flow cytometry, the suggested use of these reagents is in a final volume of 100 µL	
Other Applications	Since applications vary, you should determine the optimum working dilution for the product that is appropriate for your specific need.	

For Research Use Only. Not for Diagnostic or Therapeutic Use.

Corporate Offices: 160 Oxmoor Blvd • Birmingham, AL 35209 • USA **Mailing Address:** P.O. Box 26221 • Birmingham, AL 35260 • USA
Tel: 205.945.1774 • U.S. and Canada: 800.722.2255 • **Fax:** 205.945.8768
Email: info@southernbiotech.com • **Website:** www.southernbiotech.com

Handling and Storage

- The purified (UNLB) antibody is supplied as 0.5 mg purified immunoglobulin in 1.0 mL of borate buffered saline, pH 8.2. *No preservatives or amine-containing buffer salts added*. Store at 2-8°C.
- The fluorescein (FITC), Cyanine 5 (CY5), Alexa Fluor® 488 (AF488), and Alexa Fluor® 555 (AF555) conjugates are supplied as 0.5 mg in 1.0 mL of PBS/NaN_3. Store at 2-8°C.
- The alkaline phosphatase (AP) conjugate is supplied as 1.0 mL in a stock solution of 50 mM Tris/1 mM $MgCl_2$/50% glycerol, pH 8.0, containing NaN_3 as preservative. Store at 2-8°C or long-term at -20°C.
- The horseradish peroxidase (HRP) conjugate is supplied as 1.0 mL in a stock solution of 50% glycerol/50% PBS, pH 7.4. No preservative added. Store at 2-8°C or long-term at -20°C.
- The biotin (BIOT) conjugate is supplied as 0.5 mg in 1.0 mL of PBS/NaN_3. Store at 2-8°C.
- The R-phycoerythrin (PE) and allophycocyanin (APC) conjugates are supplied as 0.25 mg in 1.0 mL of PBS/NaN_3 and a stabilizing agent. Store at 2-8°C. **Do not freeze!**
- The Spectral Red® (SPRD) and R-phycoerythrin-Cyanine 5.5 (PE/CY5.5) conjugates are supplied as 0.25 mg in 1.0 mL of PBS/NaN_3 and a stabilizing agent. Store at 2-8°C. **Do not freeze!**
- Protect fluorochrome-conjugated forms from light. Reagents are stable for the period shown on the label if stored as directed.

Warning

Some reagents contain sodium azide. Please refer to product specific SDS.

References

1. Mansfield LS, Bell JA, Wilson DL, Murphy AJ, Elsheikha HM, Rathinam VA, et al. C57BL/6 and congenic interleukin-10-deficient mice can serve as models of Campylobacter jejuni colonization and enteritis. Infect Immun. 2007;75:1099-115. (ELISA)
2. Zhang H, Jia C, Xi H, Li S, Yang L, Wang Y. Specific inhibition of Candida albicans growth in vitro by antibodies from experimental Candida keratitis mice. Exp Eye Res. 2011;93:50-8. (ELISA)
3. Shaw CA, Galarneau J, Bowenkamp KE, Swanson KA, Palmer GA, Palladino G, et al. The role of non-viral antigens in the cotton rat model of respiratory syncytial virus vaccine-enhanced disease. Vaccine. 2013;31:306-12. (ELISPOT)
4. Xia Y, Ross GD. Generation of recombinant fragments of CD11b expressing the functional β-glucan-binding lectin site of CR3 (CD11b/CD18). J Immunol. 1999;162:7285-93. (FC)
5. Force WR, Glass AA, Benedict CA, Cheung TC, Lama J, Ware CF. Discrete signaling regions in the lymphotoxin-β receptor for tumor necrosis factor receptor-associated factor binding, subcellular localization, and activation of cell death and NF-κB pathways. J Biol Chem. 2000;275:11121-9. (FC)
6. Céfai D, Favre L, Wattendorf E, Marti A, Jaggi R, Gimmi CD. Role of Fas ligand expression in promoting escape from immune rejection in a spontaneous tumor model. Int J Cancer. 2001;91:529-37. (FC)
7. Samten B, Wizel B, Shams H, Weis SE, Klucar P, Wu S, et al. CD40 ligand trimer enhances the response of $CD8^+$ T cells to Mycobacterium tuberculosis. J Immunol. 2003;170:3180-6. (FC)
8. Ritelli M, Amadori M, Tagliabue S, Pacciarini ML. Use of a macrophage cell line for rapid detection of Mycobacterium bovis in diagnostic samples. Vet Microbiol. 2003;94:105-20. (FC)
9. McNees AL, Mahr JA, Ornelles D, Gooding LR. Postinternalization inhibition of adenovirus gene expression and infectious virus production in human T-cell lines. J Virol. 2004;78:6955-66. (FC)
10. Mongini PK, Inman JK, Han H, Kalled SL, Fattah RJ, McCormick S. Innate immunity and human B cell clonal expansion: effects on the recirculating B2 subpopulation. J Immunol. 2005;175:6143-54. (FC)
11. Kawabata A, Momoi Y, Inoue-Murayama M, Iwasaki T. Canine mdr1 gene mutation in Japan. J Vet Med Sci. 2005;67:1103-7. (FC)
12. Clancy RM, Neufing PJ, Zheng P, O'Mahony M, Nimmerjahn F, Gordon TP, et al. Impaired clearance of apoptotic cardiocytes is linked to anti-SSA/Ro and -SSB/La antibodies in the pathogenesis of congenital heart block. J Clin Invest. 2006;116:2413-22. (FC)
13. Shao DD, Suresh R, Vakil V, Gomer RH, Pilling D. Pivotal Advance: Th-1 cytokines inhibit, and Th-2 cytokines promote fibrocyte differentiation. J Leukoc Biol. 2008;83:1323-33. (FC, ICC)
14. Garnett CT, Talekar G, Mahr JA, Huang W, Zhang Y, Ornelles DA, et al. Latent species C adenoviruses in human tonsil tissues. J Virol. 2009;83:2417-28. (FC)
15. Drewniak A, van Raam BJ, Geissler J, Tool AT, Mook OR, van den Berg TK, et al. Changes in gene expression of granulocytes during in vivo granulocyte colony-stimulating factor/dexamethasone mobilization for transfusion purposes. Blood. 2009;113:5979-98. (FC)
16. Miller LA, Gerriets JE, Tyler NK, Abel K, Schelegle ES, Plopper CG, et al. Ozone and allergen exposure during postnatal development alters the frequency and airway distribution of CD25+ cells in infant rhesus monkeys. Toxicol Appl Pharmacol. 2009;236:39-48. (FC)
17. Montalvão F, Almeida GM, Silva EM, Borges VM, Vasconcellos R, Takiya CM, et al. Apoptotic lymphocytes treated with IgG from Trypanosoma cruzi infection increase TNF-α secretion and reduce parasite replication in macrophages. Eur J Immunol. 2010;40:417-25. (FC, ICC)
18. Chmielewski M, Hombach AA, Abken H. CD28 cosignalling does not affect the activation threshold in a chimeric antigen receptor-redirected T-cell attack. Gene Ther. 2011;18:62-72. (FC)
19. Fournier P, Aigner M, Schirrmacher V. Targeting of IL-2 and GM-CSF immunocytokines to a tumor vaccine leads to increased anti-tumor activity. Int J Oncol. 2011;38:1719-29. (FC)
20. Guzman R, Valente EG, Pretorius J, Pacheco E, Qi M, Bennett BD, et al. Expression of ORAI1, a plasma membrane resident subunit of the CRAC channel, in rodent and non-rodent species. J Histochem Cytochem. 2014;62:864-78. (FC)
21. Dasso JF, Obiakor H, Bach H, Anderson AO, Mage RG. A morphological and immunohistological study of the human and rabbit appendix for comparison with the avian bursa. Dev Comp Immunol. 2000;24:797-814. (IHC-FS, IHC-PS)
22. Crawford JR, Pilling D, Gomer RH. FcγRI mediates serum amyloid P inhibition of fibrocyte differentiation. J Leukoc Biol. 2012;92:699-711. (ICC)
23. Tosi MF, Zakem H. Surface expression of Fcγ receptor III (CD16) on chemoattractant-stimulated neutrophils is determined by both surface shedding and translocation from intracellular storage compartments. J Clin Invest. 1992;90:462-70. (WB)
24. Rifkin IR, Leadbetter EA, Beaudette BC, Kiani C, Monestier M, Shlomchik MJ, et al. Immune complexes present in the sera of autoimmune mice activate rheumatoid factor B cells. J Immunol. 2000;165:1626-33. (ELISA)
25. Singh N, Johnstone DB, Martin KA, Tempera I, Kaplan MJ, Denny MF. Alterations in nuclear structure promote lupus autoimmunity in a mouse model. Dis Model Mech. 2016;9:885-97. (ELISA)
26. Hande S, Notidis E, Manser T. Bcl-2 obstructs negative selection of autoreactive, hypermutated antibody V regions during memory B cell development. Immunity. 1998;8:189-98. (IHC-FS)
27. Notidis E, Hande S, Manser T. Enforced expression of Bcl-2 selectively perturbs negative selection of dual reactive antibodies. Dev Immunol. 2001;8:223-34. (IHC-FS)
28. Szijártó V, Guachalla LM, Visram ZC, Hartl K, Varga C, Mirkina I, et al. Bactericidal monoclonal antibodies specific to the lipopolysaccharide O antigen from multidrug-resistant Escherichia coli clone ST131-O25b:H4 elicit protection in mice. Antimicrob Agents Chemother. 2015;59:3109-16. (WB)
29. Ohnaga T, Shimada Y, Takata K, Obata T, Okumura T, Nagata T, et al. Capture of esophageal and breast cancer cells with polymeric microfluidic devices for CTC isolation. Mol Clin Oncol. 2016;4:599-602.

Corporate Offices: 160 Oxmoor Blvd • Birmingham, AL 35209 • USA **Mailing Address:** P.O. Box 26221 • Birmingham, AL 35260 • USA
Tel: 205.945.1774 • U.S. and Canada: 800.722.2255 • **Fax:** 205.945.8768
Email: info@southernbiotech.com • **Website:** www.southernbiotech.com