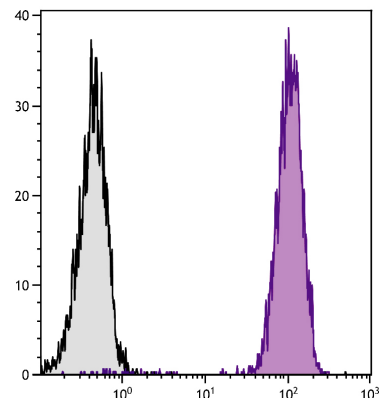




## Mouse Anti-Chicken MHC Class I

| Cat. No. | Format               | Size   |
|----------|----------------------|--------|
| 8345-01  | Purified (UNLB)      | 0.5 mg |
| 8345-02  | Fluorescein (FITC)   | 0.5 mg |
| 8345-08  | Biotin (BIOT)        | 0.5 mg |
| 8345-09  | R-phycoerythrin (PE) | 0.1 mg |



Chicken peripheral blood lymphocytes were stained with Mouse Anti-Chicken MHC Class I-UNLB (SB Cat. No. 8345-01) followed by Goat Anti-Mouse IgG1, Human ads-FITC (SB Cat. No. 1070-02).

### Overview

|                          |  |
|--------------------------|--|
| <b>Clone</b>             | F21-2  |
| <b>Isotype</b>           | Mouse (BALB/c) IgG <sub>1</sub> κ                |
| <b>Immunogen</b>         | Unknown  |
| <b>Specificity</b>       | Chicken/Turkey MHC Class I α-chain; Mr 40-43 kDa |
| <b>Alternate Name(s)</b> | B-F  |

### Description

Like their mammalian counterparts, avian MHC Class I molecules, also known as B-F antigens, consist of a highly polymorphic α-chain noncovalently bound to the invariant β<sub>2</sub>-microglobulin subunit. MHC Class I molecules are expressed on most nucleated cells where they present endogenously synthesized antigenic peptides to CD8<sup>+</sup> T lymphocytes, which are usually cytotoxic T cells. The monoclonal antibody F21-2 also reacts with turkey MHC Class I.

### Applications

FC – Quality tested <sup>3,8-12</sup>  
 IHC-FS – Reported in literature <sup>3,4</sup>  
 EM – Reported in literature <sup>13</sup>  
 IP – Reported in literature <sup>1,2,5</sup>  
 WB – Reported in literature <sup>1,2</sup>  
 Purification – Reported in literature <sup>1,2,6</sup>  
 Block – Reported in literature <sup>7</sup>

### Working Dilutions

|                           |  |                                |
|---------------------------|--|--------------------------------|
| <b>Flow Cytometry</b>     | FITC and BIOT conjugates   | ≤ 1 μg/10 <sup>6</sup> cells   |
|                           | PE conjugate   | ≤ 0.2 μg/10 <sup>6</sup> cells |
|                           | For flow cytometry, the suggested use of these reagents is in a final volume of 100 μL   |                                |
| <b>Other Applications</b> | Since applications vary, you should determine the optimum working dilution for the product that is appropriate for your specific need. |                                |

**For Research Use Only. Not for Diagnostic or Therapeutic Use.**

## Handling and Storage

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- The purified (UNLB) antibody is supplied as 0.5 mg of purified immunoglobulin in 1.0 mL of borate buffered saline, pH 8.2. *No preservatives or amine-containing buffer salts added.* Store at 2-8°C.
- The fluorescein (FITC) conjugate is supplied as 0.5 mg in 1.0 mL of PBS/NaN<sub>3</sub>. Store at 2-8°C.
- The biotin (BIOT) conjugate is supplied as 0.5 mg in 1.0 mL of PBS/NaN<sub>3</sub>. Store at 2-8°C.
- The R-phycoerythrin (PE) conjugate is supplied as 0.1 mg in 1.0 mL of PBS/NaN<sub>3</sub> and a stabilizing agent. Store at 2-8°C. **Do not freeze!**
- Protect fluorochrome-conjugated forms from light. Reagents are stable for the period shown on the label if stored as directed.

## Warning

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Some reagents contain sodium azide. Please refer to product specific SDS.

## References

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1. Salomonsen J, Skjødtt K, Crone M, Simonsen M. The chicken erythrocyte-specific MHC antigen. Characterization and purification of the B-G antigen by monoclonal antibodies. *Immunogenetics*. 1987;25:373-82. (IP, WB, Purification)
2. Møller LB, Kaufman J, Verland S, Salomonsen J, Avila D, Lambiris JD, et al. Variations in the cytoplasmic region account for the heterogeneity of the chicken MHC class I (B-F) molecules. *Immunogenetics*. 1991;34:110-20. (IP, WB, Purification)
3. Dunon D, Salomonsen J, Skjødtt K, Kaufman J, Imhof BA. Ontogenic appearance of MHC class I (B-F) antigens during chicken embryogenesis. *Dev Immunol*. 1990;1:127-35. (IHC-FS, FC)
4. Subedi K, Yoshimura Y. Expression of MHC class I and II in growing ovarian follicles of young and old laying hens, *Gallus domesticus*. *J Poult Sci*. 2005;42:101-9. (IHC-FS)
5. Dunon D, Kaufman J, Salomonsen J, Skjoedt K, Vainio O, Thiery J, et al. T cell precursor migration towards  $\beta$ 2-microglobulin is involved in thymus colonization of chicken embryos. *EMBO J*. 1990;9:3315-22. (IP, IHC-FS)
6. Wallny H, Avila D, Hunt LG, Powell TJ, Riegert P, Salomonsen J, et al. Peptide motifs of the single dominantly expressed class I molecule explain the striking MHC-determined response to Rous sarcoma virus in chickens. *Proc Natl Acad Sci USA*. 2006;103:1434-9. (Purification)
7. Haghighi HR, Read LR, Haeryfar SM, Behboudi S, Sharif S. Identification of a dual-specific T cell epitope of the hemagglutinin antigen of an h5 avian influenza virus in chickens. *PLoS One*. 2009;4(11):e7772. (Block)
8. Del Cacho E, Gallego M, Lillehoj HS, López-Bernard F, Sánchez-Acedo C. Avian follicular and interdigitating dendritic cells: isolation and morphologic, phenotypic, and functional analyses. *Vet Immunol Immunopathol*. 2009;129:66-75. (FC)
9. Singh S, Briles WE, Lupiani B, Collisson EW. Avian influenza viral nucleocapsid and hemagglutinin proteins induce chicken CD8<sup>+</sup> memory T lymphocytes. *Virology*. 2010;399:231-8. (FC)
10. Sunkara LT, Achanta M, Schreiber NB, Bommineni YR, Dai G, Jiang W, et al. Butyrate enhances disease resistance of chickens by inducing antimicrobial host defense peptide gene expression. *PLoS One*. 2011;6(11):e27225. (FC)
11. Verweij MC, Lipińska AD, Koppers-Lalic D, van Leeuwen WF, Cohen JI, Kinchington PR, et al. The capacity of UL49.5 proteins to inhibit TAP is widely distributed among members of the genus *Varicellovirus*. *J Virol*. 2011;85:2351-63. (FC)
12. Meyerhoff RR, Ali RA, Liu K, Huang G, Koci MD. Comprehensive analysis of commercially available mouse antichickens monoclonal antibodies for cross-reactivity with peripheral blood leukocytes from commercial turkeys. *Poult Sci*. 2012;91:383-92. (FC, Turkey Reactivity)
13. del Cacho E, Gallego M, Lee SH, Lillehoj HS, Quilez J, Lillehoj EP, et al. Induction of protective immunity against *Eimeria tenella* infection using antigen-loaded dendritic cells (DC) and DC-derived exosomes. *Vaccine*. 2011;29:3818-25. (EM)